

Patient demographics
 Patient demographics (age, time elapsed since onset of symptoms) are available for the 257 samples used in the study. The table below shows the positive results broken down by age of the patient:

Age	LumiraDx SARS-CoV-2 Ag (n = 81)		
	Total #	Positive	Prevalence
≤ 5 years	13	0	N/A
6 to 21 years	29	6	20.7%
22 to 59 years	200	70	35.0%
≥ 60 years	15	5	33.3%

Days since symptom onset	Cumulative RT-PCR Positive(+)	Cumulative LumiraDx Positive(+)	PPA	95% Confidence interval	
0	6	6	100.0%	61.0%	100.0%
1	12	12	100.0%	75.8%	100.0%
2	28	28	100.0%	87.9%	100.0%
3	37	37	100.0%	90.6%	100.0%
4	55	54	98.2%	90.4%	99.7%
5	61	60	98.4%	91.3%	99.7%
6	67	66	98.5%	92.0%	99.7%
7	73	72	98.6%	92.6%	99.8%
8	75	74	98.7%	92.8%	99.8%
9	75	74	98.7%	92.8%	99.8%
10	77	76	98.7%	93.0%	99.8%
11	80	79	98.8%	93.3%	99.8%
12	83	81	97.6%	91.6%	99.3%

Reference RT-PCR Assay						95% Wilson Score CI	
						LCI	UCI
LumiraDx SARS-CoV-2 Ag Test		POS	NEG	Total	PPA	97.6%	99.3%
	POS	81	6	87	NPA	96.6%	98.4%
	NEG	2	168	170	Prevalence	32.3%	38.2%
	TOTAL	83	174	257			

PPA - Positive Percent Agreement (Sensitivity)
 NPA - Negative Percent Agreement (Specificity)
 UCI - Upper Confidence Interval
 CI - Confidence Interval
 LCI - Lower Confidence Interval

Clinical Performance - Nasopharyngeal Swabs
 The performance of the SARS-CoV-2 Ag Test was established with 255 nasopharyngeal swabs prospectively collected from individual subjects between August 2020 and September 2020 during the 2020 COVID pandemic. Subjects were presenting with symptoms of COVID-19 being screened for infection. Samples were collected from 6 sites across the United States. Swabs were collected and extracted into the LumiraDx Extraction Buffer. Samples were tested fresh within 1h of collection and tested according to the Product Insert. The performance of the LumiraDx SARS-CoV-2 Ag Test was compared to the results from nasopharyngeal samples collected into 3ml universal transport medium (UTM) and tested with an EUA authorized PCR method.

Age	LumiraDx SARS-CoV-2 Ag (n = 39)		
	Total #	Positive	Prevalence
≤ 5 years	22	0	0.0%
6 to 21 years	59	9	15.3%
22 to 59 years	150	28	18.7%
≥ 60 years	24	2	8.3%

Days since symptom onset	Cumulative PCR Positive (+)	LumiraDx Positive (+)	PPA	LCI	UCI	NPA	LCI	UCI
0	2	2	100.0%	34.2%	100.0%	100.0%	75.8%	100.0%
1	6	6	100.0%	61.0%	100.0%	100.0%	93.4%	100.0%
2	9	9	100.0%	70.1%	100.0%	100.0%	96.2%	100.0%
3	17	17	100.0%	81.6%	100.0%	98.6%	94.9%	99.6%
4	22	22	100.0%	85.1%	100.0%	98.8%	95.7%	99.7%
5	23	23	100.0%	85.7%	100.0%	98.4%	95.3%	99.4%
6	26	26	100.0%	87.1%	100.0%	98.5%	95.6%	99.5%
7	34	34	100.0%	89.8%	100.0%	98.5%	95.7%	99.5%
8	36	36	100.0%	90.4%	100.0%	98.6%	95.8%	99.5%
9	36	36	100.0%	90.4%	100.0%	98.6%	95.9%	99.5%
10	39	38	97.4%	86.8%	99.5%	98.1%	95.2%	99.3%
11		39	97.5%	87.1%	99.6%	97.7%	94.6%	99.0%
12	40	39	97.5%	87.1%	99.6%	97.7%	94.7%	99.0%

Reference RT-PCR Assay					95% Wilson Score CI		
					LCI	UCI	
LumiraDx SARS-CoV-2 Ag Test		POS	NEG	Total	PPA	97.5%	99.6%
	POS	39	5	44	NPA	97.7%	99.0%
	NEG	1	210	211	Prevalence	15.7%	20.7%
	TOTAL	40	215	255			

NIH study on three Lateral Flow OTC rapid Antigen tests (Not including LumiraDx)
 A prospective clinical study was conducted between January 2021 and May 2022 as a component of the Rapid Acceleration of Diagnostics (RADx) initiative from the National Institutes of Health (NIH). A total of 7,361 individuals were enrolled via a decentralized clinical study design, with a broad geographical representation of the United States. Per inclusion criteria, all individuals were asymptomatic upon enrollment in the study and at least 14 days prior to it and did not have a SARS-CoV-2 infection in the three months prior to enrollment. Participants were assigned to one of three EUA authorized SARS-CoV-2 OTC rapid antigen tests to conduct serial testing (every 48 hours) for 15 days. If an antigen test was positive, the serial-antigen testing result is considered positive. At each rapid antigen testing time point, study subjects also collected a nasal swab for comparator testing using a home collection kit (using a 15-minute normalization window between swabs). SARS-CoV-2 infection status was determined by a composite comparator method on the day of the first antigen test, using at least two highly sensitive EUA RT-PCRs. If results of the first two molecular test were discordant a third highly sensitive EUA RT-PCR test was performed, and the final test result was based upon the majority rule. Study participants reported symptom status throughout the study using the MyDataHelps app. Two-day serial antigen testing is defined as performing two antigen tests 36 - 48 hours apart. Three-day serial antigen testing is defined as performing three antigen tests over five days with at least 48 hours between each test. Out of the 7,361 participants enrolled in the study, 5,609 were eligible for analysis. Among eligible participants, 154 tested positive for SARS-CoV-2 infection based on RT-PCR, of which 97 (62%) were asymptomatic on the first day of their infection, whereas 57 (39%) reported symptoms on the first day of infection. Pre-symptomatic subjects were included in the positive percent agreement (PPA) of asymptomatic individuals. If they were asymptomatic on the first day of antigen testing, regardless of whether they developed symptoms at any time after the first day of testing. Performance of the antigen test with serial testing in individuals is described in the table below. **Note: This study was not performed using LumiraDx assay.** Data establishing PPA of COVID-19 antigen serial testing compared to the molecular comparator single day testing throughout the course of infection with serial testing. Data is from all antigen tests in the study combined (LumiraDx not included in this study), however, the data is being applied to all antigen tests to support serial testing.

Note: This study was not performed using Lumiraux assay.

Data establishing PPA of COVID-19 antigen serial testing compared to the molecular comparator single day testing throughout the course of infection with serial testing. Data is from all antigen tests in the study combined (LumiraDx not included in this study). However, the data is being applied to all antigen tests to support serial testing.

Days after first PCR Positive Test Result	Asymptomatic on first day of testing			Symptomatic on first day of testing		
	Ag Positive / PCR Positive (Antigen Test Performance % PPA)					
	1 Test	2 Tests	3 Tests	1 Test	2 Tests	3 Tests
0	9/97 (9.3%)	35/89 (39.3%)	44/78 (56.4%)	34/57 (59.6%)	47/51 (92.2%)	44/47 (93.6%)
2	17/34 (50.0%)	23/34 (67.6%)	25/32 (78.1%)	58/62 (93.5%)	59/60 (98.3%)	43/43 (100%)
4	16/21 (76.2%)	15/20 (75.0%)	13/15 (86.7%)	55/58 (94.8%)	53/54 (98.1%)	39/40 (97.5%)
6	20/28 (71.4%)	21/27 (77.8%)	16/18 (88.9%)	27/34 (79.4%)	26/33 (78.8%)	22/27 (81.5%)
8	13/23 (56.5%)	13/22 (59.1%)	4/11 (36.4%)	12/17 (70.6%)	12/17 (70.6%)	7/11 (63.6%)
10	5/9 (55.6%)	5/8 (62.5%)	-	4/9 (44.4%)	3/7 (42.9%)	-

1 Test = one (1) test performed on the noted days after first PCR positive test result. Day 0 is the first day of documented infection with SARS-CoV-2.
 2 Tests = two (2) tests performed on average of 48 hours apart. The first test performed on the indicated day and the second test performed 48 hours later.
 3 Tests = three (3) tests performance on average of 48 hours apart. The first test performed on the indicated day, the second test performed 48 hours later, and a final test performed 48 hours after the second test.

Analytical performance
Limit of Detection - LOD (analytical sensitivity)
 Limit of Detection (LoD) studies determined the lowest detectable concentration of SARS-CoV-2 at which 100% of all (true positive) replicates test positive. The LoD for the LumiraDx SARS-CoV-2 Ag Test was established using limiting dilutions of gamma-irradiated SARS-CoV-2 (BEIResources NR-52287). The NR-52287 is a preparation of SARS-Related Coronavirus 2 (SARS-CoV2), isolate USA WAI/2020, that has been inactivated by gamma-irradiation at 5 x 10⁶ RADs. The material was supplied frozen at a concentration of 2.8 x 10⁶ TCID₅₀/mL. **Limit of Detection (LoD) screening**
 An initial LoD screening study was performed using a 5-fold serial dilutions (six dilutions in total) of the gamma-irradiated virus made in pooled negative human nasal matrix starting at a test concentration of 2 x 10⁶ TCID₅₀/mL (as shown in table below) and processed for each study as described above. These dilutions were tested in triplicate. The volume of solution spiked onto each swab was 50µL. The lowest concentration at which all (3 out of 3 replicates) were positive was chosen for LoD Range finding. This was 32 TCID₅₀/mL. Based upon the testing procedure for this study the LoD of 32 TCID₅₀/mL equates to 1.6 TCID₅₀/swab.

SARS-CoV-2 tested (TCID ₅₀ /mL)	Test result
20000	3/3 positive
4000	3/3 positive
800	3/3 positive
160	3/3 positive
32	3/3 positive
6.4	0/3 positive

SARS-CoV-2 tested (TCID ₅₀ /mL)		Test result	
32		3/3 positive	
16		0/3 positive	
8		1/3 positive	
4		0/3 positive	

Limit of Detection (LoD) confirmation
 The LoD of the LumiraDx SARS-CoV-2 Ag Test was then confirmed by testing 20 replicates with concentrations at the tentative Limit of Detection. The final LoD of the LumiraDx SARS-CoV-2 Ag Test was determined to be the lowest concentration resulting in positive detection of twenty (20) out of twenty (20) replicates. Based on this testing the LoD for nasal swab samples was confirmed as: 32 TCID₅₀/mL. Based upon the testing procedure for this study the LoD of 32 TCID₅₀/mL equates to 1.6 TCID₅₀/swab.

Starting Material Concentration	Estimated LOD	No. Positive/Total	% Positive
2.8 x 10 ⁶ TCID ₅₀ /mL	32 TCID ₅₀ /mL	20/20	100

Omnicron Pool 2 - Live Dilution	Assay #1	Assay #2	LumiraDx SARS-CoV-2 Ag Test*
CH-N2 Ave.	Percent Positive (n=5)	Percent Positive (n=5)	Percent Positive (n=5)
Dilution 1	19.8	100	100
Dilution 2	20.8	100	100
Dilution 3	21.5	100	100
Dilution 4	22.7	100	100
Dilution 5	23.6	100	100
Dilution 6	24.0	60	40
Dilution 7	24.8	0	20
Dilution 8	25.8	0	0
Dilution 9	27.4	0	0
Dilution 10	28.1	0	0
Dilution 11	29.1	0	0

*Testing was conducted using a multiplexed version of the test, which comprises the same SARS-CoV-2 test design and components.

Microorganism	Source	Concentration	Cross-Reactivity (Yes/No)	Interference (Yes/No)
Human coronavirus 229E	Zeptomatrix	1 x 10 ⁶ PFU/mL	No (3/3 negative)	No (3/3 positive)
Human coronavirus OC43	Zeptomatrix	1 x 10 ⁶ PFU/mL	No (3/3 negative)	No (19/20 positive)
Human coronavirus NL63	Zeptomatrix	9.87 x 10 ³ PFU/mL	No (3/3 negative)	No (3/3 positive)
MERS coronavirus	Zeptomatrix	7930 PFU/mL	No (2/2 negative)	No (3/3 positive)
Adenovirus (e.g. C1 Ad. 71)	Zeptomatrix	1 x 10 ⁶ PFU/mL	No (3/3 negative)	No (3/3 positive)
Human Metapneumovirus (hMPV)	Zeptomatrix	1 x 10 ⁶ PFU/mL	No (3/3 negative)	No (3/3 positive)
Parainfluenza virus Type 1	Zeptomatrix	1 x 10 ⁶ PFU/mL	No (3/3 negative)	No (3/3 positive)
Parainfluenza virus Type 2	Zeptomatrix	1 x 10 ⁶ PFU/mL	No (3/3 negative)	No (3/3 positive)
Parainfluenza virus Type 3	Zeptomatrix	1 x 10 ⁶ PFU/mL	No (3/3 negative)	No (3/3 positive)
Parainfluenza virus Type 4a	Zeptomatrix	1 x 10 ⁶ PFU/mL	No (3/3 negative)	No (3/3 positive)
Influenza A H3N2 (Wisconsin/67/05)	Zeptomatrix	8.82 x 10 ⁴ PFU/mL	No (3/3 negative)	No (3/3 positive)
Influenza A H1N1	Zeptomatrix	1 x 10 ⁶ PFU/mL	No (3/3 negative)	No (3/3 positive)
Influenza B (Malaysia/2506/04)	Zeptomatrix	2.92 x 10 ⁴ PFU/mL	No (3/3 negative)	No (19/20 positive)
Enterovirus	Zeptomatrix	1 x 10 ⁶ PFU/mL	No (3/3 negative)	No (3/3 positive)
Respiratory syncytial virus	Zeptomatrix	1 x 10 ⁶ PFU/mL	No (3/3 negative)	No (3/3 positive)
Rhinovirus	Zeptomatrix	4.17 x 10 ⁶ PFU/mL	No (3/3 negative)	No (3/3 positive)
Haemophilus influenzae	Zeptomatrix	1 x 10 ⁶ CFU/mL	No (3/3 negative)	No (3/3 positive)
Streptococcus pneumoniae	Zeptomatrix	1 x 10 ⁶ CFU/mL	No (3/3 negative)	No (3/3 positive)
Streptococcus pyogenes	Zeptomatrix	1 x 10 ⁶ CFU/mL	No (3/3 negative)	No (3/3 positive)
Candida albicans	Zeptomatrix	1 x 10 ⁶ CFU/mL	No (3/3 negative)	No (3/3 positive)
Pooled human nasal wash	LumiraDx	14% v/v	No (3/3 negative)	No (3/3 positive)
Bordetella pertussis	Zeptomatrix	1 x 10 ⁶ CFU/mL	No (3/3 negative)	No (3/3 positive)
Mycoplasma pneumoniae	ATCC	1 x 10 ⁶ CFU/mL	No (3/3 negative)	No (3/3 positive)
Chlamydia pneumoniae	ATCC	1 x 10 ⁶ CFU/mL	No (3/3 negative)	No (3/3 positive)
Legionella pneumophila	Zeptomatrix	1 x 10 ⁶ CFU/mL	No (3/3 negative)	No (3/3 positive)
Mycobacterium tuberculosis	Zeptomatrix	1 x 10 ⁶ CFU/mL	No (3/3 negative)	No (3/3 positive)
Pneumocystis jirovecii	Zeptomatrix	1 x 10 ⁶ CFU/mL	No (3/3 negative)	No (3/3 positive)
Pseudomonas Aeruginosa	Zeptomatrix	1 x 10 ⁶ CFU/mL	No (3/3 negative)	No (3/3 positive)
Staphylococcus Epidermidis	Zeptomatrix	1 x 10 ⁶ CFU/mL	No (3/3 negative)	No (3/3 positive)
Streptococcus Salivarius	Zeptomatrix	1 x 10 ⁶ CFU/mL	No (3/3 negative)	No (3/3 positive)
Staphylococcus aureus	ATCC	1 x 10 ⁶ CFU/mL	No (3/3 negative)	No (3/3 positive)

To estimate the likelihood of cross-reactivity with SARS-CoV-2 of organisms that were not available for wet testing, *in silico* analysis using the Basic Local Alignment Search Tool (BLAST) managed by the National Center for Biotechnology Information (NCBI) was used to assess the degree of protein sequence homology.

- For Human Coronavirus HKU1, homology exists between the SARS-CoV-2 nucleocapsid protein and Human Coronavirus HKU1. BLAST results showed 30 sequence IDs, all nucleocapsid protein, showing homology. Sequence ID AGW27840.1 had the highest alignment score and was found to be 39.1% homologous across 76% of the sequences, this is relatively low but cross-reactivity cannot be fully ruled out.
- For SARS-Coronavirus, high homology exists between the SARS-CoV-2 nucleocapsid protein and SARS-Coronavirus. BLAST results showed 68 sequence IDs, mostly nucleocapsid protein, showing homology. Sequence ID AAR87518.1, had the highest alignment score isolated from a human patient and was found to be 90.76% homologous across 100% of the sequence. This is high and cross-reactivity is likely.
- For MERS-Coronavirus, high homology exists between the SARS-CoV-2 nucleocapsid protein and MERS-Coronavirus. BLAST results showed at least 114 sequence IDs, mostly nucleocapsid protein, showing homology. Sequence IDs AHY61344.1 and AWH65950.1, had the highest alignment scores isolated from a human patient and were found to be 49.4% and 50.3% homologous across 88% of the sequence. Whilst this potentially represents moderate cross-reactivity testing of the MERS virus at 7930 PFU/mL showed no reactivity (see table above).

Endogenous interference studies
 A study was performed to demonstrate that potentially interfering substances that may be found in the upper respiratory tract in symptomatic subjects (including over the counter medications) do not interfere with detection of SARS-CoV-2 in the LumiraDx SARS-CoV-2 Ag Test. Each substance was tested in triplicate in the absence or presence of SARS-CoV-2 at 3 x LoD. Substances for testing were selected based on the respiratory samples guidance in http://www.accessdata.fda.gov/cdrh_docs/reviews/K112177.pdf. The final concentration of the substances tested are documented in the table below.

Interfering substance	Concentration	Interference (Yes/No)
Benzocaine	150 mg/dL	No (3/3 Negative, 3/3 Positive)
Blood (human)	5%	No (3/3 Negative, 3/3 Positive)
Mucin	5 mg/mL	No (3/3 Negative, 3/3 Positive)
Naso GEL (NeilMed)	5% v/v	No (3/3 Negative, 3/3 Positive)
CVS Nasal Drops (phenylephrine)	15% v/v	No (3/3 Negative, 3/3 Positive)
Afrin (Oxymetazoline)	15% v/v	No (3/3 Negative, 3/3 Positive)
CVS Nasal Spray (Cromolyn)	15% v/v	No (3/3 Negative, 3/3 Positive)
Zicam Cold Remedy	5% v/v	No (3/3 Negative, 3/3 Positive)
Homeopathic (Alkaloi)	10% v/v	No (3/3 Negative, 3/3 Positive)
Sore Throat Phenol Spray	15% v/v	No (3/3 Negative, 3/3 Positive)
Tobramycin	3.3 mg/dL	No (3/3 Negative, 3/3 Positive)
Mupirocin	0.15 mg/dL	No (3/3 Negative, 3/3 Positive)
Fluticasone	0.000126 mg/dL	No (5/5 Negative, 4/4 Positive)
Tamiflu (Osetamivir phosphate)	500 mg/dL	No (3/3 Negative, 3/3 Positive)
Budesonide	0.00063 mg/dL	No (3/3 Negative, 3/3 Positive)
Biotin	0.35 mg/dL	No (3/3 Negative, 3/3 Positive)

Interfering substance	Concentration	Interference (Yes/No)
Methanol	150 mg/dL	No (19/20 Negative, 3/3 Positive)
Acetylsalicylic Acid	3 mg/dL	No (3/3 Negative, 3/3 Positive)
Diphenhydramine	0.0774 mg/dL	No (3/3 Negative, 3/3 Positive)
Dextromethorphan	0.00156 mg/dL	No (19/20 Negative, 3/3 Positive)
Dexamethasone	1.2 mg/dL	No (3/3 Negative, 3/3 Positive)
Mucinex	5%	No (3/3 Negative, 3/3 Positive)

High dose hook effect
 High Dose Hook Effect studies determine the level at which false negative results can be seen when very high levels of target are present in a tested sample. To determine if the LumiraDx SARS-CoV-2 Ag Test suffers from any high dose hook effect, increasing concentrations of gamma-irradiated SARS-CoV-2 virus (BEI Resources NR-52287) were tested up to a concentration of 1.4 x 10⁶ TCID₅₀/mL. In this study, the starting material was spiked into a volume of pooled human nasal matrix obtained from healthy donors and confirmed negative for SARS-CoV-2. At each dilution, 50 µL samples were added to swabs and the swabs processed for testing on the LumiraDx SARS-CoV-2 Ag Test as per the Product Insert using the procedure appropriate for patient nasal swab samples. No impact on test performance or high dose hook effect was observed up to 1.4 x 10⁶ TCID₅₀/mL of gamma-irradiated SARS-CoV-2 with the LumiraDx SARS-CoV-2 Ag Test.

Test dilution	Concentration (TCID ₅₀ /mL)	Mean signal (ADC Units)
1	0	495
2	62.5	26100.6
3	250	63013.8
4	1000	83451.8
5	1.4 x 10 ⁶	86220

Point of care use
 The LumiraDx SARS-CoV-2 Ag Test was used by 8 untrained users in 4 sites across the United States. Untrained users tested 132 patients and ran 148 tests.

References

1.

World Health Organisation www.who.int

2.

Centers for Disease Control and Prevention www.cdc.gov

Symbols glossary



Temperature limitation



Manufacturer



In Vitro Diagnostic Medical Device



Catalogue Number



Batch code/Lot Number



Indicates a medical device that has been sterilized using ethylene oxide



Indicates that a medical device that should not be used if the package has been damaged or opened and that the user should consult the instructions for use for additional information



Use by



Consult Instructions for Use



Do Not Re-use



Prescription Use Only



Do not re-sterilize



Indicates the presence of the Radio Frequency Identification (RFID) reader/tag.



Date of manufacture