

# dynamic BIOSENSORS heliX plus Reducing Agent Kit 1 User **Manual**

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dynamic BIOSENSORS heliX plus Reducing Agent Kit 1



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#### Tube

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- · Allows for reduction of disulfide bonds using TCEP.
- Compatible with proteins and antibodies (MW > 20 kDa).
- Site specific reduction of antibodies.
- Immediate coupling of reduced molecules is possible (refer to user manual HK-MAL).
- Reduction of multiple ligands can be performed at the same time.
- Includes reagents for two individual reduction reactions.

# **Product Description**

Order Number: RK-PA-1

Table 1. Contents and Storage Information

Material	Сар	Amount	Storage
TCEP (500 mM)	Red	2 x 20 μL	-20°C
Reducing Buffer [1]	White	2 x 1.8 mL	-20°C
Purification spin column	Red	4 x	2-8°C
2.0 mL reaction tubes for purification spin column		4 x	RT
Centrifugal filter unit (10 k Da MWCO)		2 x	RT
Centrifugation collection t ube		4 x	RT

For research use only.

This product has a limited shelf life, please see expiry date on label.

IMPORTANT	Products may be shipped at different temperatures, but storage should a dhere to the guidelines outlined in the Table. The resin slurry in the purific ation spin column contains 0.02 % sodium azido
	ation spin column contains 0.02 % sodium azide.

# **Additional Materials Required**

Table 2. Additional Materials

Material	Comments
Benchtop microcentrifuge	Required speed range of between 1,000 x g to 13,000 x g
Vortex	
1.5 mL reaction tubes	

All necessary solutions and buffers are included in the kit.

# 3-Step Reduction of a Biomolecule in a Reaction Tube

Please read the entire protocol before starting and perform all steps without interruption.

TI	IP	This protocol can be performed simultaneously for multiple coupling reactions
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## **Buffer Exchange of Protein/Antibody**

- 1. Wet the centrifugal filter unit membrane with 100 μL Reducing Buffer.
- 2. Add approx. 200  $\mu$ g (up to 500  $\mu$ g) of protein/antibody to the filter unit from step 1. If necessary add Reducing Buffer to the filter until a maximum volume of 450  $\mu$ L is reached and centrifuge at 13,000 x g (up to 14,000 x g) for 5 min and discard flow-through.
- 3. Add 350 µL of Reducing Buffer and centrifuge at 13,000 x g for 5 min and discard flow-through again.
- 4. Repeat step 3, one more time with a final centrifugation time of 10 min to get a final volume of approx. 80 μL.
- 5. Collect sample either with a 100  $\mu$ L pipette into a new tube or by placing the filter device upside down in a new centrifugal tube, centrifuge for 2 min at 1000 x g.

## **Reducing with TCEP**

- 1. Prepare a 2.5 mM TCEP solution in Reducing Buffer by adding 1  $\mu$ L of 500 mM TCEP into 199  $\mu$ L Reducing Buffer.
  - Prepare always directly before use.
- 2. Add Reducing Buffer to the sample vial for a total volume of 98 μL and add 2 μL of 2.5 mM TCEP solution to obtain a final concentration of 50 μM TCEP. Mix the reaction by pipetting up and down.
- 3. Incubate the solution at 37°C at 400 rpm for 2 h to ensure complete reduction.

# **Purification and Concentration**

- 1. Equilibrate two purification spin columns (red cap) for one TCEP wash out:
  - a. Remove the column's bottom seal and loosen cap (do not remove cap).
  - **b**. Place the column in a 2.0 mL reaction tube.
  - **c**. Centrifuge at  $1,500 \times g$  for 1 min to remove the storage solution.
  - **d.** Add 400 μL of Reducing Buffer to the column's resin bed. Centrifuge at 1,500 × g for 1 min to remove buffer.
  - **e**. Repeat step d and discard the resulting buffer from the reaction tube. The purification spin column should now be in a dry state.

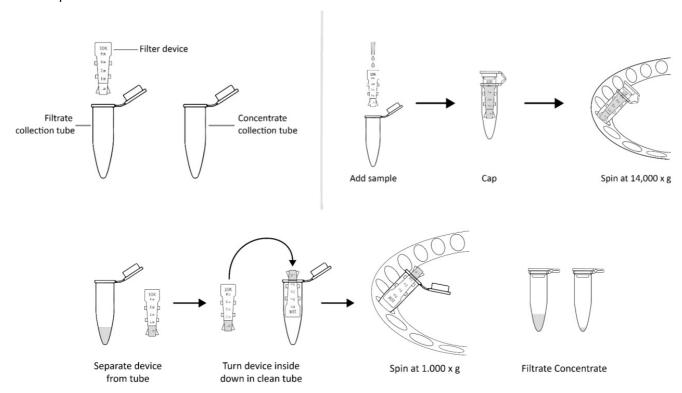
#### **TCEP** wash

- a. Place the columns from the previous step in new 1.5 mL reaction tubes.
- **b**. Remove the cap of spin column number 1 and pour delicately the sample to the top of the resin bed.
- **c**. Centrifuge at 1,500 x g for 2 min to collect the sample (flow-through). Discard the purification spin column after use.
- **d**. Remove the cap of spin column number 2 and pour the sample from the previous step to the resin bed.
- **e**. Centrifuge at 1,500 x g for 2 min to collect the sample (flow-through). Discard the purification spin column after use.
- f. Thiol coupling according user manual (HK-MAL-1 or HK-MAL-2) of reduced sample can be performed.

#### **Additional Information**

# **Buffer Exchange and Concentration with Centrifugal Filter Units**

- 1. Take one centrifugal filter unit, add the appropriate volume of buffer in the filter device, and cap it.
- 2. Place capped filter device into the centrifuge rotor, aligning the cap strap toward the center of the rotor; counterbalance with a similar device.
- 3. Spin the device at 13,000 x g (or 14,000 x g) for the given time.
- 4. Remove the flow through and repeat steps 1-3.
- 5. Remove the assembled device from the centrifuge and separate the filter device from the microcentrifuge tube.
- 6. To recover the conjugate, place the filter device upside down in a clean centrifugal tube, aligning open cap towards the center of the rotor; counterbalance with a similar device. Spin for 2 minutes at 1,000 x g to transfer the sample from the device to the tube.



#### **Useful Order Numbers**

Table 3. Order Numbers

Product Name	Amount	Order No
heliX ® Thiol coupling kit 1	5 x	HK-MAL-1
heliX ® Thiol coupling kit 2	5 x	HK-MAL-2
Centrifugal filter unit (3 kDa MWCO	5 pcs.	CF-003-5
Centrifugal filter unit (10 kDa MWC O)	5 pcs.	CF-010-5
10x Buffer A [2]	50 mL (yielding 500 mL)	BU-P-150-10
5x Buffer B [3]	50 mL (yielding 250 mL)	BU-P-1000-5

#### Contact

Dynamic Biosensors GmbH	Dynamic Biosensors, Inc	
Perchtinger Str. 8/10	300 Trade Center, Suite 1400	
81379 Munich	Woburn, MA 01801	
Germany	USA	

Order Information <u>order@dynamic-biosensors.com</u> Technical Support <u>support@dynamic-biosensors.com</u>

- 1. Reducing Buffer: 100 mM Na2HPO4/NaH2PO4, 300 mM NaCl, 5 mM EDTA, pH 7.4
- 2. Buffer A: 50 mM Na2HPO4/NaH2PO4, 150 mM NaCl, pH 7.2
- 3. Buffer B: 50 mM Na2HPO4/NaH2PO4, 1 M NaCl, pH 7.2

#### **CUSTOMER SUPPORT**

#### www.dynamic-biosensors.com

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#### **Documents / Resources**



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# References

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